



HAL
open science

Brain and gonadal aromatase as potential targets of endocrine disrupting chemicals in a model species, the zebrafish (*Danio rerio*)

Nathalie Hinfray, Olivier Palluel, Cyril Turies, C. Cousin, Jean-Marc Porcher, François Brion

► To cite this version:

Nathalie Hinfray, Olivier Palluel, Cyril Turies, C. Cousin, Jean-Marc Porcher, et al.. Brain and gonadal aromatase as potential targets of endocrine disrupting chemicals in a model species, the zebrafish (*Danio rerio*). *Environmental Toxicology*, 2006, 21 (4), pp.332-337. 10.1002/tox.20203 . ineris-00961901

HAL Id: ineris-00961901

<https://ineris.hal.science/ineris-00961901>

Submitted on 20 Mar 2014

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Brain and gonadal aromatase as potential targets of endocrine disrupting chemicals in a model species, the zebrafish (*Danio rerio*).

Hinfray N., Palluel O., Turies C., Cousin C., Porcher J.M., Brion F*.

Unité d'évaluation des risques écotoxicologiques, Direction des Risques Chroniques, Institut National de l'Environnement Industriel et des Risques (INERIS), BP 2, F-60550 Verneuil-en-Halatte, France

† Corresponding author. Tel.: +33 (0)3 44 55 65 12; Fax: +33 (0)3 44 55 67 67

E-mail address: Francois.Brion@ineris.fr

ABSTRACT

Many chemicals in the aquatic environment are able to adversely affect *in vitro* brain and ovarian aromatase expression/activity. However, it remains to be determined if these substances elicit *in vivo* effect in fish. With the view to further understanding possible effects of endocrine disrupting chemicals (EDCs) on aromatase function, we first developed methods to measure brain and ovarian aromatase expression/activity in a model species, the zebrafish, and assessed the effect of estradiol (E2) and androstatrienedione (ATD), a steroidal aromatase inhibitor. We showed that CYP19b gene was predominantly expressed in the brain whereas in the ovary CYP19a mRNA level was predominant. Moreover, aromatase activities (AA) were higher in brain than in ovary. In adult zebrafish, E2 treatment had no effect on aromatase expression/activity in brain, whereas at larval stage, E2 strongly triggered CYP19b expression. In the ovaries, E2 led to a complete inhibition of both CYP19a expression and AA. Exposure to ATD led to a total inhibition of both brain and ovarian AA but had no effect on CYP19 transcripts abundance. Together, these results provide relevant knowledge concerning the characterization of aromatase in the zebrafish, and reinforce the idea that brain and ovarian aromatase are promising markers of EDCs in fish and deserve further *in vivo* studies.

Keywords : zebrafish, aromatase, brain, gonad, biomarker.

INTRODUCTION

To date, most attention on endocrine disrupting chemicals (EDCs) has been focused on compounds that interact with the estrogen receptor (ER). However, the endocrine system may also be disrupted by environmental substances through pathways and mechanisms others than those that are ER-mediated. Function of the hypothalamic-pituitary-gonadal (HPG) axis can be affected by xenobiotics that affect metabolism of sex steroid hormones. In this regard the biosynthesis of steroid hormones provides enzymatic targets for EDCs, particularly the steps catalysed by cytochrome P450-dependent enzymes (Sanderson and Van den Berg, 2003). Among these enzymes, cytochrome P450 aromatase (CYP19) is a crucial steroidogenic enzyme catalysing the final, rate-limiting step in the conversion of androgens into estrogens (Simpson *et al.*, 1994). Recent studies reported alteration of steroidogenesis associated with adverse reproductive effects in wild fish collected from contaminated sites (Noaksson *et al.*, 2001; Orlando *et al.*, 2002; Lavado *et al.*, 2004). However the nature (and the levels) of substances involved in these biological responses remains to be determined. In the laboratory, different substances from diverse chemical family have been shown to disrupt vertebrate aromatase (Monteiro *et al.*, 2000; Sanderson *et al.*, 2002). In fish, however, most of the data were obtained from *in vitro* studies (Monod *et al.*, 1993) and little is known about their *in vivo* effects.

The purpose of this study was to assess the effect of the natural steroidal estrogen, estradiol (E2), product of the aromatisation reaction, and androstratienedione (ATD), a steroidal aromatase inhibitor, on aromatase expression and activities in the zebrafish (*Danio rerio*). The zebrafish is a prominent vertebrate model in a variety of biological discipline (Hill *et al.*, 2005) and is extensively used for assessing the effects of estrogenic compounds at various biological levels of organization (Brion *et al.*, 2002, 2004; Fenske and Segner, 2004; Nash *et al.*, 2004). In the zebrafish, two distinct aromatase genes have been isolated (CYP19a and

CYP19b) and their 5'-flanking region characterised (Kazeto *et al.*, 2001). While their tissue and ontogenic expression profiles are relatively well described (Trant *et al.*, 2001), data on brain and ovarian AA in control and exposed-zebrafish are missing. With the view to further understanding possible effects of EDCs on aromatase function, detailed information on gene expression and enzyme activities in both the gonads and brain are needed and this was the first aim of this study.

MATERIAL AND METHODS

Fish exposure to E2 and ATD

Wild type zebrafish (*Danio rerio*) were obtained from our breeding unit. Female zebrafish were exposed for 7 days to 17- β -estradiol (E2, 10 nM), 1,4,6-androstatrien-3,17-dione (ATD, 1 μ M) or solvent (DMSO). Exposures were realised under semi-static conditions with a total renewal of the water every day. Each substance was tested in duplicate separated tanks. One replicate was used to measure aromatase activity (N = 10 fish), and the other one to measure CYP19 mRNA levels (N = 7 fish).

Zebrafish larvae of 17 days post fertilisation (dpf) were exposed to either E2 (10 nM) alone or in combination with ICI 182-780 (Tocris, USA) or to solvent alone (DMSO) for 72 hours. Each experimental group was constituted of 20 zebrafish larvae exposed in 100 ml of water. One half of the water was renewed every day.

Fish dissection and tissue sampling

Adult fish were euthanized by an overdose of MS-222, measured and weighted. Brain and gonads were removed, weighted and the gonado somatic index (GSI) was calculated.

Tissues (or larvae) used for subsequent determination of mRNA levels were immediately stored at 4°C in a solution of RNAlaterTM (Sigma-Aldrich, France). Samples were kept at 4°C overnight and conserved at -20°C until measurement. For AA measurement, tissues were rinsed in ice cold KCl (0.15 M) and homogenised in a 50 mM potassium phosphate buffer, pH 7.4, containing 1 mM PMSF, 1 mM EDTA and 20% glycerol (v/v) in a ratio of 1:2 (w/v). S9 fractions were isolated by centrifugation of the homogenates (10 000 g, 15 min, 4°C). For the brains, S9 were aliquoted and stored at -80°C until used while for the ovaries, supernatants

were subsequently ultra-centrifuged at 100,000 *g* (90 min, 4°C). AA were measured in ovarian microsomes instead of S9 fractions to avoid variation of protein content during the reproductive cycle which may lead to incorrect measurement of AA in this tissue. The microsomal pellets were resuspended in the same buffer as used for the homogenisation. The total amount of proteins was determined by the method of Bradford with BSA (Bovine Serum Albumin, Sigma-Aldrich, France) as standard (Bradford, 1976).

Determination of CYP19a and CYP19b mRNA levels

CYP19 mRNA levels were measured by a branched DNA assay (QuantiGene, Genospectra, USA). It directly measures RNA and does not require reverse transcription and cDNA amplification. The bDNA assays were performed according to the manufacturer instructions. Tissues (or 10 pooled whole-body zebrafish larvae) were lysed and incubated in a 96-well plate coated with synthetic oligonucleotide in the presence of a specific probe set designed according to the CYP19a and CYP19b mRNA sequences (gene bank accession number AF183906 and AF183908 respectively). Capture probe allowed capture of the target mRNA to the synthetic oligonucleotide. Blocking Probe linearised the target mRNA and a label probe hybridised to the target mRNA and to a branched DNA (bDNA) coupled with Alkaline-Phosphatase-bound Probes. Addition of a chemiluminescence substrate (Dioxetan) yields a luminescence signal that is proportional to the amount of mRNA present in the sample. Quantification of luminescence was made on a microplate luminometer (Victor², Perkin Elmer, France). CYP19 expression values were normalised to a housekeeping gene, zf β -actin (gene bank accession number NM 131031). Measurement of target and housekeeping genes were realised in duplicate.

Aromatase assay

Brain and ovarian AA were measured in individual fish by the tritiated water assay modified from Thompson and Siiteri (1974). The following additions were made in a final volume of 500 μL : 200 μg of microsomal proteins (ovary) or S9 fraction proteins (brain), 20 μM of NADPH, 1 mM of NADP, 10 mM of G6P, 2 U/ml of G6PDH, 50 mM potassium phosphate buffer, pH 7.4. The mixture was preincubated at 27 °C for 10 min, and the reaction started with the addition of 150 nM of [1β - ^3H (N)]androst-4-ene-3,17-dione (Perkin Elmer, France). The reaction was stopped after 1 h of incubation by adding 1 ml of chloroform. The aqueous layer was extracted twice with chloroform and once with charcoal (5%, w/v). 150 μL of the aqueous phase was mixed with 750 μL of OptiPhase 'Hi safe' 3 (Perkin Elmer, France) before scintillation counting (Microbeta, Perkin Elmer, France). AA measurements were realised in duplicates. Results were expressed in fmol/mg/min.

Data analysis and statistics

Differences between groups were analysed for statistical significance with the Student's t-test. Results are expressed as means \pm standard deviation, and differences between groups were considered to be significant if $p < 0.05$.

RESULTS

Baseline aromatase expression and activity in adult female zebrafish

The transcript abundance for CYP19a and CYP19b genes is shown in the Fig. 1A. In the brain, levels of CYP19b mRNA expression was about 45-fold higher than CYP19a expression, while, in the ovaries, levels of CYP19a mRNA expression was about 3-fold higher than CYP19b expression. Moreover, the level of expression of CYP19b was higher in the brain than the expression of CYP19a in the ovary. At enzymatic level, AA measured in female zebrafish by the tritiated water assay were higher in brain than in ovaries (Fig. 1B). There was about a 4-fold difference between brain and ovarian AA (34.0 ± 25.3 and 8.1 ± 8.2 fmol/mg/min respectively).

Effect of exposure to E2 and ATD

In adult female zebrafish, E2 exposure had no effect on CYP19b expression in the brain but significantly inhibited CYP19a expression in the ovary (Fig. 2A). At enzymatic level, brain AA were not affected, but ovarian AA were totally inhibited (Fig. 2B).

In zebrafish larvae exposed between 17 and 20 dpf to E2, there was no significant effect on CYP19a expression. However, a 16-fold increase in CYP19b expression was measured (Fig. 3). In addition, co-treatment with the pure antagonist of the ER receptor ICI 182-780 led to a significant inhibition of the E2-induced CYP19b mRNA levels.

In ATD-exposed fish, there were no significant effects neither on the transcript abundance of CYP19b gene in the brain nor on CYP19a expression in the ovary (Fig. 4A). However, ATD totally inhibited both brain and ovarian AA (Fig. 4B).

DISCUSSION

CYP19 genes expression and AA in adult female zebrafish

In contrast to mammals, many teleosts fish possess two forms of the aromatase gene in their genome, as it was demonstrated for zebrafish (Kishida and Callard, 2001). CYP19a and CYP19b transcripts, as measured by the bDNA assays, were both detected in ovarian and brain tissues of adult female zebrafish which is in agreement with previously reported CYP19 genes expression measured by RT-PCR (Trant *et al.*, 2001; Fenske and Segner, 2004). In the ovary, CYP19a mRNA level was higher compared to CYP19b, whereas CYP19b expression was predominant in the brain. Cerebral CYP19b was shown to be expressed at particularly high levels in adult female (about 600 fold CYP19a expression in the ovary) which is well in accordance with the difference observed by Trant *et al.* (2001). At the cellular level, ovarian CYP19a expression was localised in granulosa cells surrounding follicles (Goto-Kazeto *et al.*, 2004); in the brain, P450aromataseB (AroB) was found to be express exclusively in radial glial cells at high levels in males and females (Menuet *et al.*, 2005). Teleosts fish are well known for their exceptionally high cerebral levels of aromatase (Pasmanik and Callard, 1988; Callard *et al.*, 2001). However, the functional outcomes of elevated aromatase expression in the brain of teleosts fish are still unresolved. Since teleosts fish shows a continuous neurogenesis throughout life, one hypothesis is that the high levels of neuroestrogens synthesis in adult may be related to the remarkable neuroplasticity and regenerative potential of the adult fish central nervous system (Callard *et al.*, 2001; Forlano *et al.*, 2001).

Further characterisation was achieved by measuring brain and ovarian AA in female zebrafish. In agreement with the CYP19 transcripts abundance, we showed that brain AA were significantly higher than ovarian AA which is in accordance with the general pattern of

AA described in other teleosts fish (Pasmanik and Callard, 1988; Gonzalez and Piferrer, 2002). These differences can be accounted for differences in the levels of mRNA expression in the two tissues (Trant *et al.*, 2001; our study) and/or be due to the higher catalytic activity (V_{max}) of the brain aromatase compared to the ovarian aromatase (data not shown).

Effect of exposure of zebrafish E2 and ATD on CYP19 expression and AA

At adult stage, E2 had no significant effect on CYP19b expression and AA in the brain of female zebrafish. In contrast, short-term exposure of zebrafish larvae to E2 strongly up-regulated CYP19b expression. This result confirms previous data obtained by RT-PCR on RNA from total zebrafish embryos and larvae exposed to E2 (Kishida and Callard, 2001). *In toto* hybridisation and immunohistochemistry experiments further revealed that E2 causes strong expression of AroB messengers and proteins in radial glial cells of zebrafish embryos and larvae (Menuet *et al.*, 2005). Additionally, the E2 dependant induction of the CYP19b gene was blocked by co-treatment with an excess of the pure ER antagonist ICI 182-780 indicating that functional ERs were involved. In teleosts fish brain, CYP19b gene is known to be under the control of a positive autoregulatory feedback loop driven by E2, the product of aromatization (Callard *et al.*, 2001). On a molecular basis, it was recently demonstrated that the E2-dependent regulation involves a direct transcriptional action of ERs requiring the synergistic effect of ERE and $\frac{1}{2}$ ERE in the promoter region of the CYP19b gene (Menuet *et al.*, 2005). The absence of effect of E2 on aromatase in adult zebrafish brain is not surprising since the endogenous estrogenic stimulation on aromatase expression/activities is already high due to the positive autoregulatory feedback loop driven by E2 (Callard *et al.*, 2001). Our results clearly demonstrated that the CYP19b expression is very sensitive to (xeno)-estrogen in zebrafish larvae in comparison with adult fish and reinforce the idea that AroB is a

promising marker of estrogenic compounds in zebrafish early life stages (Menuet *et al.*, 2005). In contrast to the dramatic alteration of the CYP19b gene expression, E2 had no effect on CYP19a expression in zebrafish larvae. In zebrafish fry exposed to high concentration of EE2, conflicting results were obtained since RT-PCR analysis revealed no effect or down-regulation of the CYP19a gene (Trant *et al.*, 2001; Kazeto *et al.*, 2004). At adult stage, both CYP19a expression and AA were totally inhibited in the ovary of E2-exposed fish and our results suggest that the inhibitory effect of E2 on AA in ovary of mature female is mediated through a transcriptional inhibition of the CYP19a gene. However, the exact mechanism of action of E2 on CYP19a expression remains to be determined. It may be attributed to a negative feedback action of E2 on gonadotropins release, which are known to stimulate aromatase gene expression and activities in ovarian follicles (Gen *et al.*, 2001; Kagawa *et al.*, 2003). However, a direct effect of E2 on the ovary cannot be excluded. Indeed, it has been demonstrated that the inhibitory effect of E2 on steroidogenic enzymes mRNA levels in undifferentiated testis of rainbow trout did not imply follicle stimulating hormone (Baron *et al.*, 2005). The absence of ERE in the promoter region of CYP19a gene do not support an ER-mediated effect. It is interesting to note that similar concentration of E2 leads to an alteration of oogenesis in females, the ovaries of exposed fish being composed mainly of immature oocytes (Brion *et al.*, 2000). It can be suggested that the effects seen at histological level are mediated, at least in part, by inhibition of ovarian aromatase.

Exposure of female zebrafish to the steroidal aromatase inhibitor ATD resulted in dramatic inhibition of AA in both brain and ovary. ATD is known to inhibit vertebrate aromatase by binding irreversibly to the active site of the enzyme (Yue and Brodie, 1997). We previously showed that ATD exhibited a high efficiency at inhibiting *in vitro* brain and ovarian microsomal AA in rainbow trout (Hinfray *et al.*, 2004). The *in vivo* effects of ATD on AA are thus consistent with the *in vitro* inhibitory effect of this molecule. In contrast to the

effect seen at the enzymatic level, exposure to ATD had no effect neither on the transcriptional activity of the CYP19b gene nor the CYP19a gene. As previously stated, CYP19b transcription in fish brain is up-regulated by E2 through a positive feedback loop (Callard *et al.*, 2001). Thus, it could be expected that the local deprivation of E2 due to aromatase inhibition in the brain could lead to a decreasing transcript abundance of the CYP19b gene. The absence of effect might be due to the short-term exposure duration of fish to ATD and it is likely that a prolonged exposure time would result in a significant effect, as shown in letrozole-exposed zebrafish larvae for 30 days (Kazeto *et al.*, 2004).

CONCLUSION

This study provides relevant data on aromatase expression at the gene and enzymatic levels in brain and in ovary of adult female fish. We showed that CYP19b expression was predominant in brain whereas CYP19a expression was predominant in ovary and that AA were higher in brain than in ovary. Our results indicate that E2 and ATD exposures deeply affect aromatase expression and/or activities in the zebrafish. The observed effects were dependent on the sub-cellular level at which aromatase function was assessed (gene expression, AA), the target tissue (brain, ovary) and the life stage of development (larvae, adult). From these results it appears that measurements of both CYP19 gene expression and AA in the zebrafish are relevant and promising molecular and biochemical markers of EDCs in fish.

ACKNOWLEDGEMENTS

This study was supported by the French Ministry of Ecology and Sustainable Development (Budget Civil de Recherche et Développement, BCRD Grant AP17-02). N. Hinfray is funded by INERIS and ARNT (National association for technical research).

REFERENCES

Baron, D., Fostier, A., Breton, B., Guiguen, Y., 2005. Androgen and estrogen treatments alter steady state messengers RNA (mRNA) levels of testicular steroidogenic enzymes in the rainbow trout, *Oncorhynchus mykiss*. *Mol Reprod Dev*, 71 (4): 471-479.

Bradford, M., 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem*, 72: 248-254.

Brion F., Triffault, G., Palazzi, X., Forraz, M., Garric, J., Migeon, B., Noury, P., Porcher, J.M., Thybaud, E., Tyler, C.R., Flammarion, P., 2000. in: Proceedings of the Third World Congress of the SETAC, 21–25 May 2000, Brighton, UK.

Brion, F., Nilsen, B.M., Eidem, J.K., Goksoyr, A., Porcher, J.M., 2002. Development and validation of an enzyme-linked immunosorbent assay to measure vitellogenin in the zebrafish (*Danio rerio*). *Environ Toxicol Chem*, 21 (8): 1699-1708.

Brion, F., Tyler, C.R., Palazzi, X., Laillet, B., Porcher, J.M., Garric, J., Flammarion, P., 2004. Impacts of 17[β]-estradiol, including environmentally relevant concentrations, on reproduction after exposure during embryo-larval-, juvenile- and adult-life stages in zebrafish (*Danio rerio*). *Aquat Toxicol*, 68 (3): 193-217.

Callard, G.V., Tchoudakova, A.V., Kishida, M., Wood, E., 2001. Differential tissue distribution, developmental programming, estrogen regulation and promoter characteristics of *cyp19* genes in teleost fish. *J Steroid Biochem Mol Biol*, 79 (1-5): 305-314.

Fenske, M., Segner, H., 2004. Aromatase modulation alters gonadal differentiation in developing zebrafish (*Danio rerio*). *Aquat Toxicol*, 67: 105-126.

Forlano, P.M., Deitcher, D.L., Myers, D.A., Bass, A.H., 2001. Anatomical distribution and cellular basis for high levels of aromatase activity in the brain of teleost fish: aromatase enzyme and mRNA expression identify glia as source. *J Neurosci*, 21 (22): 8943-8955.

Gen, K., Okuzawa, K., Kumakura, N., Yamaguchi, S., Kagawa, H., 2001. Correlation between messenger RNA expression of cytochrome P450 aromatase and its enzyme activity during oocyte development in the red seabream (*Pagrus major*). *Biol Reprod*, 65 (4): 1186-1194.

Gonzalez, A., Piferrer, F., 2002. Characterization of aromatase activity in the sea bass: effects of temperature and different catalytic properties of brain and ovarian homogenates and microsomes. *J Exp Zoolog*, 293 (5): 500-510.

Goto-Kazeto, R., Kight, K.E., Zohar, Y., Place, A.R., Trant, J.M., 2004. Localization and expression of aromatase mRNA in adult zebrafish. *Gen Comp Endocrinol*, 139 (1): 72-84.

Hill, A.J., Teraoka, H., Heideman, W., Peterson, R.E., 2005. Zebrafish as a model vertebrate for investigating chemical toxicity. *Toxicol Sci*, 86 (1): 6-19.

Hinfray, N., Porcher, J.M., Brion, F., 2004. In vitro effect of various environmental pollutants on brain and ovarian aromatase activity in rainbow trout (*Oncorhynchus mykiss*). in:

Proceedings of the 14th Congress of the SETAC Europe, 18–22 April 2004, Prague, Czech Republic.

Kagawa, H., Gen, K., Okuzawa, K., Tanaka, H., 2003. Effects of Luteinizing Hormone and Follicle-Stimulating Hormone and Insulin-Like Growth Factor-I on Aromatase Activity and P450 Aromatase Gene Expression in the Ovarian Follicles of Red Seabream, *Pagrus major*. *Biol Reprod*, 68 (5): 1562-1568.

Kazeto, Y., Ijiri, S., Place, A.R., Zohar, Y., Trant, J.M., 2001. The 5'-flanking regions of CYP19A1 and CYP19A2 in zebrafish. *Biochem Biophys Res Commun*, 288 (3): 503-508.

Kazeto, Y., Place, A.R., Trant, J.M., 2004. Effects of endocrine disrupting chemicals on the expression of CYP19 genes in zebrafish (*Danio rerio*) juveniles. *Aquat Toxicol*, 69 (1): 25-34.

Kishida, M., Callard, G.V., 2001. Distinct cytochrome P450 aromatase isoforms in zebrafish (*Danio rerio*) brain and ovary are differentially programmed and estrogen regulated during early development. *Endocrinology*, 142 (2): 740-750.

Lavado, R., Thibaut, R., Raldua, D., Martin, R., Porte, C., 2004. First evidence of endocrine disruption in feral carp from the Ebro River. *Toxicol Appl Pharmacol*, 196 (2): 247-257.

Menuet, A., Pellegrini, E., Brion, F., Gueguen, M.M., Anglade, I., Pakdel, F., Kah, O., 2005. Expression and estrogen-dependent regulation of the zebrafish brain aromatase gene. *J Comp Neurol*, 485 (4): 304-320.

Monod, G., De Mones, A., Fostier, A., 1993. Inhibition of ovarian microsomal aromatase and follicular estradiol suppression by imidazole fungicides in rainbow trout. *Mar Environ Res*, 35: 153-157.

Monteiro, P.R.R., Reis-Henriques, M.A., Coimbra, J., 2000. Polycyclic aromatic hydrocarbons inhibit in vitro ovarian steroidogenesis in the flounder (*Platichthys flesus L.*). *Aquat Toxicol*, 48 (4): 549-559.

Nash, J.P., Kime, D.E., Van der Ven, L.T., Wester, P.W., Brion, F., Maack, G., Stahlschmidt-Allner, P., Tyler, C.R., 2004. Long-term exposure to environmental concentrations of the pharmaceutical ethynylestradiol causes reproductive failure in fish. *Environ Health Perspect*, 112 (17): 1725-1733.

Noaksson, E., Tjarnlund, U., Bosveld, A.T.C., Balk, L., 2001. Evidence for Endocrine Disruption in Perch (*Perca fluviatilis*) and Roach (*Rutilus rutilus*) in a Remote Swedish Lake in the Vicinity of a Public Refuse Dump. *Toxicol Appl Pharmacol*, 174 (2): 160-176.

Orlando, E.F., Davis, W.P., Guillette, L.J., Jr., 2002. Aromatase activity in the ovary and brain of the eastern mosquitofish (*Gambusia holbrooki*) exposed to paper mill effluent. *Environ Health Perspect*, 110 Suppl 3: 429-433.

Pasmanik, M., Callard, G.V., 1988. Changes in brain aromatase and 5 alpha-reductase activities correlate significantly with seasonal reproductive cycles in goldfish (*Carassius auratus*). *Endocrinology*, 122: 1349-1356.

Sanderson, J.T., Boerma, J., Lansbergen, G.W.A., van den Berg, M., 2002. Induction and Inhibition of Aromatase (CYP19) Activity by Various Classes of Pesticides in H295R Human Adrenocortical Carcinoma Cells. *Toxicol Appl Pharmacol*, 182 (1): 44-54.

Sanderson, T., Van den Berg, M., 2003. Interactions of xenobiotics with the steroid hormone biosynthesis pathway. *Pure Appl Chem*, 75 (11-12): 1957-1971.

Simpson, E.R., Mahendroo, M.S., Means, G.D., Kilgore, M.W., Hinshelwood, M.M., Graham-Lorence, S., Amarneh, B., Ito, Y., Fisher, C.R., Michael, M.D., et al., 1994. Aromatase cytochrome P450, the enzyme responsible for estrogen biosynthesis. *Endocr Rev*, 15 (3): 342-355.

Thompson, E.A., Jr., Siiteri, P.K., 1974. Utilization of oxygen and reduced nicotinamide adenine dinucleotide phosphate by human placental microsomes during aromatization of androstenedione. *J Biol Chem*, 249 (17): 5364-5372.

Trant, J.M., Gavasso, S., Ackers, J., Chung, B.C., Place, A.R., 2001. Developmental expression of cytochrome P450 aromatase genes (CYP19a and CYP19b) in zebrafish fry (*Danio rerio*). *J Exp Zool*, 290 (5): 475-483.

Yue, W., Brodie, A.M.H., 1997. Mechanisms of the actions of aromatase inhibitors 4-hydroxyandrostenedione, fadrozole, and aminoglutethimide on aromatase in JEG-3 cell culture. *J Steroid Biochem Mol Biol*, 63 (4-6): 317-328.

FIGURE LEGENDS

Fig. 1 : Relative transcript abundance of CYP19a and CYP19b gene (normalised to beta-actin transcript abundance in the same samples) (A) and aromatase activity (B) measured in brain and ovary of adult female zebrafish. Results are expressed as means and SD. Different letters indicate statistically different values (Student's t-test, $p < 0.05$)

Fig. 2 : Effect of E2 on CYP19a and CYP19b in ovary and brain respectively (A) and on aromatase activity (B). Results are expressed as means and SD. SC = solvent control. Different letters indicate statistically different values (Student's t-test, $p < 0.05$).

Fig. 3 : Effect of E2 alone or in combination with ICI 182-780 on CYP19a and CYP19b transcript abundance in zebrafish larvae after a 72 hours *in vivo* exposure between 17 and 20 dpf. SC = solvent control. N = 2 replicate for each group tested (one replicate correspond to 10 larvae). Different letters indicate statistically different values (Student's t-test, $p < 0.05$).

Fig. 4 : Effect of ATD on CYP19a and CYP19b transcript abundance in ovary and brain respectively (A) and on aromatase activity (B). Results are expressed as means and SD. SC = solvent control. Different letters indicate statistically different values (Student's t-test, $p < 0.05$).

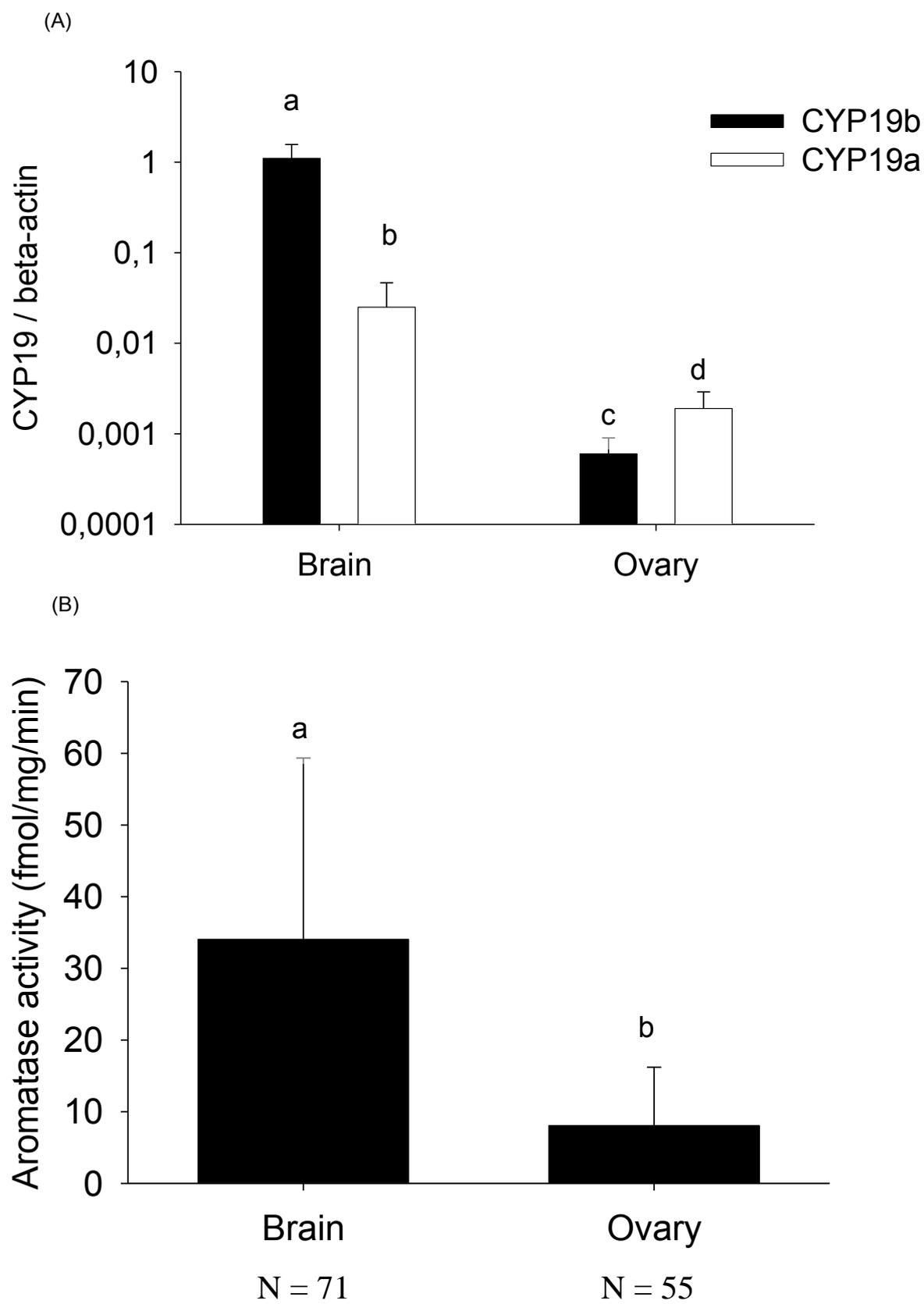


Fig. 1.

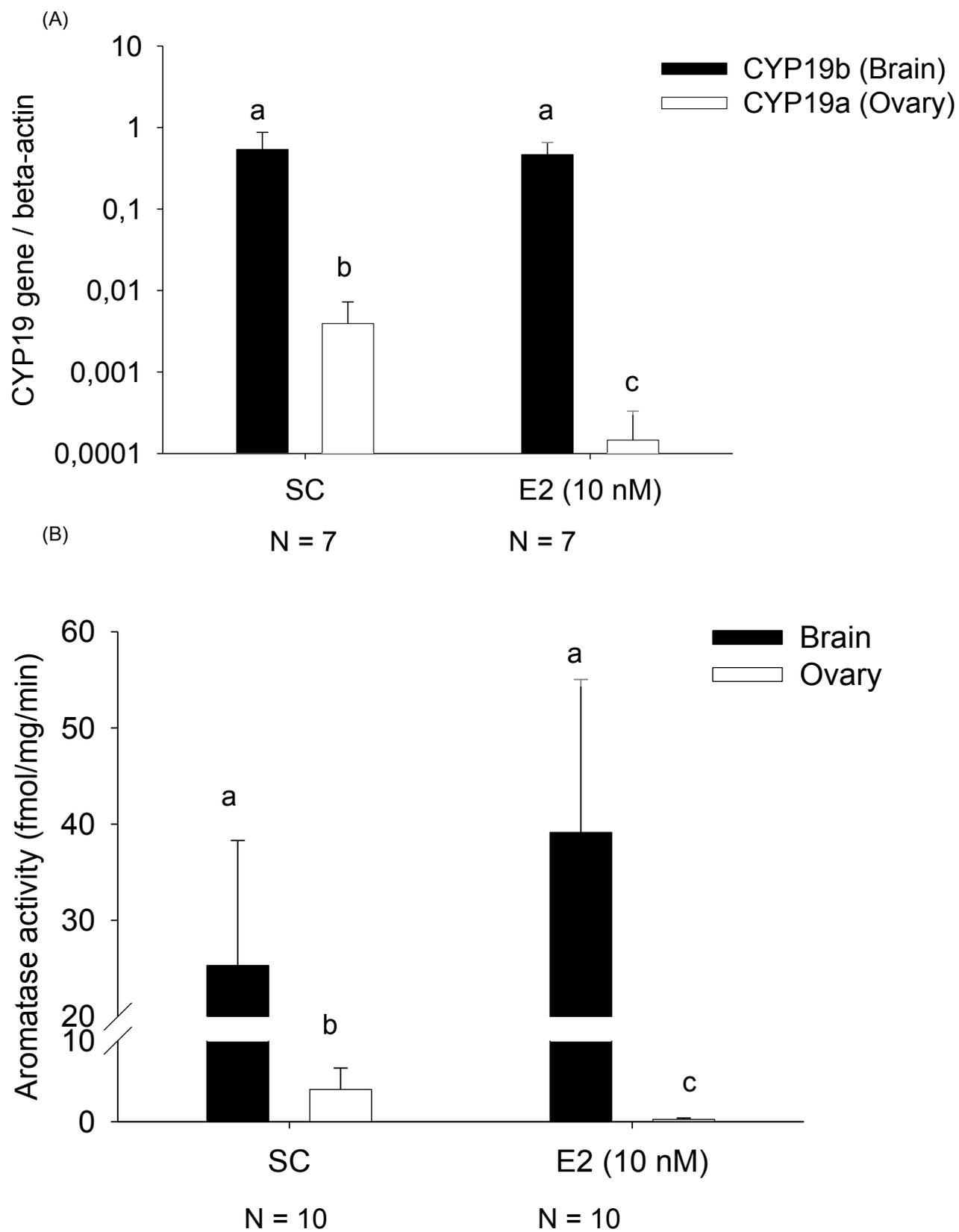


Fig. 2.

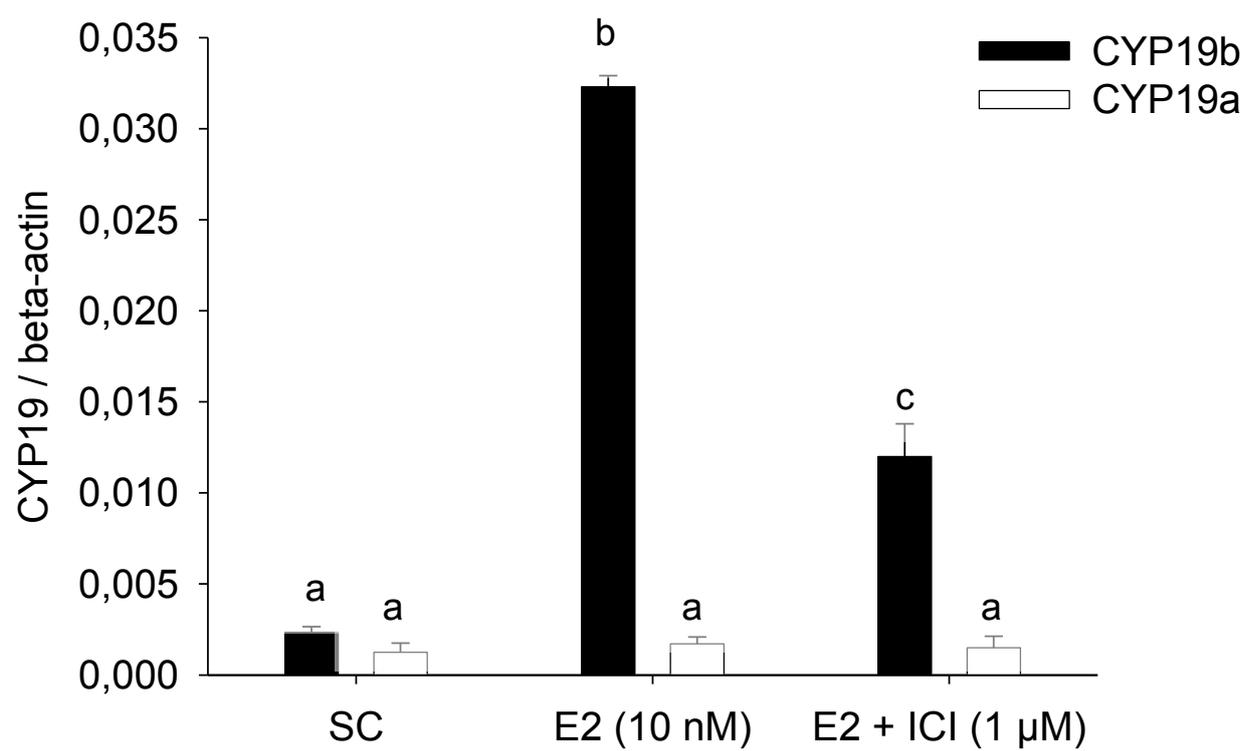


Fig. 3.

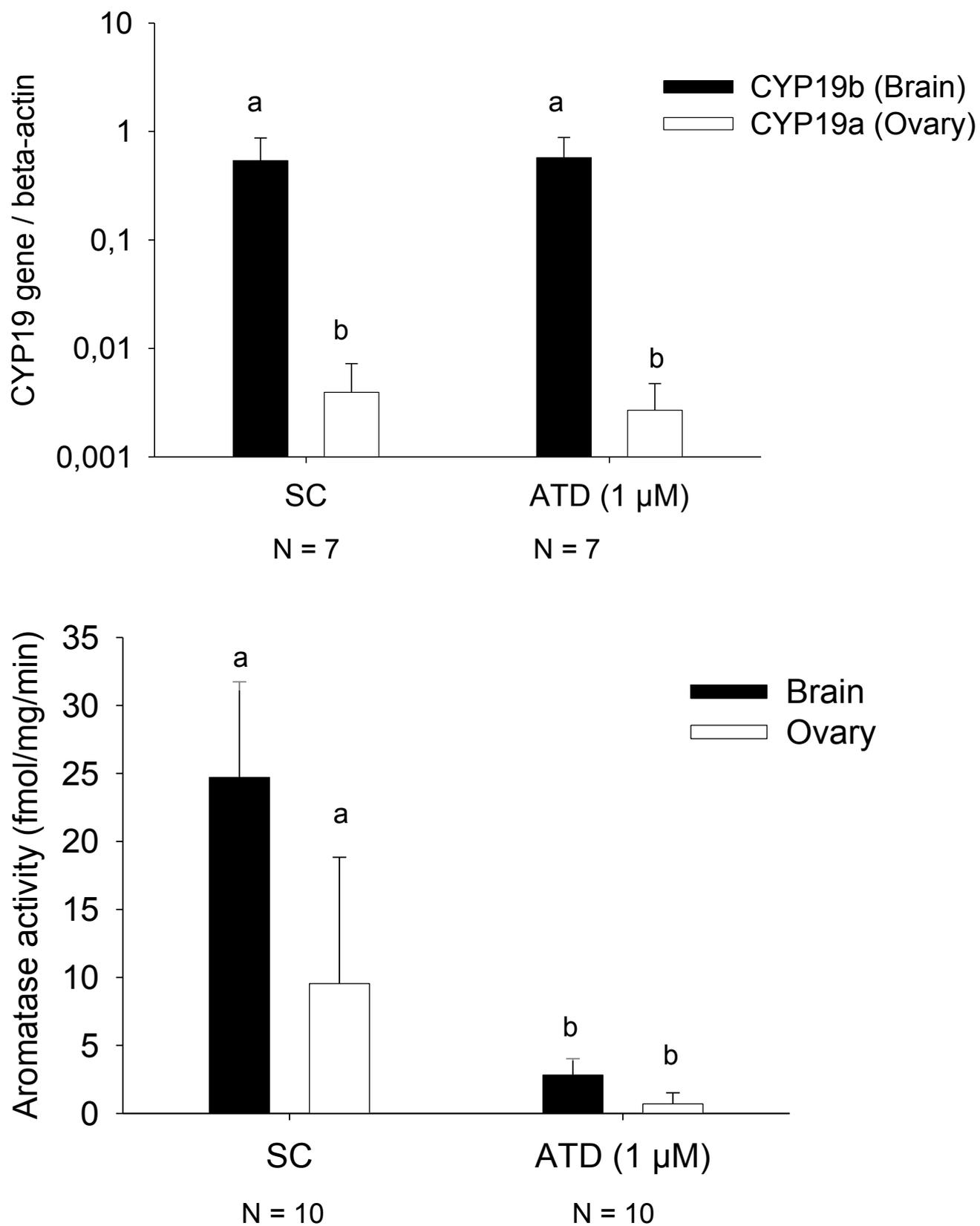


Fig. 4.